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Review

Comparison of 5-Fluoro-2'-deoxyuridine with 5-Fluorouracil and their Role in the Treatment of Colorectal Cancer

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Despite more than 30 years of intensive studies on new drugs against advanced colorectal cancer, the fluoropyrimidines remain the drugs of choice for systemic treatment and for hepatic artery infusion (HAI). This overview describes new developments in advanced colorectal cancer chemotherapy, providing a rationale for more effective use of the fluoropyrimidines, with biochemical modulation, scheduling or by revealing biochemical mechanisms of action that correlate with antitumour activity. In human colorectal cancer cell lines and various animal tumour model systems 5-fluoro-2'-deoxyuridine (FdUrd) is more effective than 5-fluorouracil (5-FU). Comparably, FdUrd's modulation by leucovorin (LV) is more potent than 5-FU. In animal studies it is shown that intermittent high-bolus administration of FdUrd generates better antitumour activity, compared with equal toxic doses or any other schedule of 5-FU. These effects are related to prolonged thymidylate synthase (TS) inhibition and the prevention of TS induction, rather than RNA incorporation. Preclinical studies with modulators such as N-phosphonacetyl-L-aspartate (PALA), WR-2721, mitomycin C and platinum derivatives provide a rationale for clinical use in the future. The first choice systemic chemotherapy of patients with advanced colorectal cancer remains 5-FU combined with LV. Some improvement in therapeutic efficacy has been achieved with locoregional HAI. In randomised studies HAI FdUrd improves the quality of life and survival as compared with optimal systemic therapy. Chronomodulation decreases toxicity, allowing dose intensification, while modulators such as LV or dexamethasone increase survival of patients treated with HAI FdUrd to 86% after 1 year. In conclusion, the clinical use of FdUrd has not been fully explored. Intermittent high-dose FdUrd, chronomodulation together with the use of modulators or drugs focused on prolonged TS inhibition, should be studied in large randomised studies. © 1998 Elsevier Science Ltd. All rights reserved.

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INTRODUCTION

COLORECTAL CANCER is the third leading cause of death from all cancers in both sexes in Europe [1]. Patients can present with liver metastases as the first or only sign (as found in 20% of all deceased patients) of advanced disease [2, 3]. Treat-

ment of metastatic disease frequently involves chemotherapy which can be given systemically or regionally. The fluoropyrimidines, 5-fluorouracil (5-FU) and 5-fluoro-2'-deoxyuridine (FdUrd) [4,5] (Figure 1) are the first choice for the treatment of patients with advanced colorectal cancer [3,6–9]. With regional hepatic artery infusion (HAI), liver metastases are almost exclusively perfused. This administration is more effective than portal vein infusion which only supplies hepatocytes. Drugs with high hepatic extraction such as the fluoropyrimidines cause minimal systemic toxicity, allowing the

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administration of higher and more effective locoregional concentrations.

5-FU with its modulator leucovorin (LV) is regarded as standard systemic chemotherapy for patients with advanced colorectal cancer [3, 7, 9-11]. FdUrd is the deoxyribonucleoside derivative of 5-FU. While it partly acts as a prodrug of 5-FU, there are many other subtle chemical differences. Its mechanism of action (Figure 2) is thought to be due to the 5fluoro-2'-deoxyuridine-5'-monophosphate (FdUMP)-mediated inhibition of thymidylate synthase (TS). 5-FU is cheaper with less systemic toxicity, so usage of FdUrd has been restricted to HAI or infusions using chronotherapy. FdUrd and 5-FU differ in their preclinical antitumour activity, clinical pharmacological characteristics and toxicity patterns [3, 6, 12]. To optimise the potential of FdUrd, chronomodulation and addition of modulators in the HAI setting were tested [3, 13, 14]. This overview compares FdUrd with 5-FU in terms of mechanisms of action, preclinical antitumour activity and application in the clinic. Using appropriate administration schedules, FdUrd showed significantly better antitumour activity in animal models compared with 5-FU. We conclude that recent insights into mechanisms of action of FdUrd suggest that the use of this drug is not optimal in the clinic.

UPTAKE AND METABOLISM OF FdUrd AND 5-FU

Both fluoropyrimidines require cellular uptake and metabolism to active products before exerting cellular effects. Ultimately, four metabolic effects of fluoropyrimidines can be distinguished: (1) inhibition of the key enzyme of *de novo* pyrimidine synthesis, TS, by the formation of FdUMP, and subsequent interference with DNA synthesis, (2) incorporation of 5-fluorouridine-5'-triphosphate (FUTP) into RNA thereby interfering with RNA synthesis, (3) incorporation of 5-fluoro-2'-deoxyuridine-5'-triphosphate (FdUTP) into DNA, and (4) the formation of 5-FU sugar derivatives [15].

The predominant metabolic pathway varies according to the fluoropyrimidine used and is determined by levels of the various intracellular enzymes for anabolism and catabolism together with the availability of competing normal substrates and cofactors. This availability varies according to the drug administration schedule as well as the tissue studied. The key mechanisms causing antiproliferative effects are believed to be TS inhibition and RNA incorporation [16–29]. The antitumour activity of 5-FU is postulated to be mainly through inhibition of TS especially when modulated by leucovorin (LV), whereas side-effects are related to RNA incorporation [17, 23, 29]. Furthermore, the mechanism of action may be a function of the schedule of drug administration; TS inhibition occurs with protracted infusion and RNA incorpora-

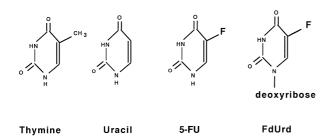


Figure 1. Chemical structures of uracil, thymine, 5-fluorouracil (5-FU) and 5-fluoro-2'-deoxyuridine (FdUrd).

tion occurs with i.v. bolus. The reversal by uridine and uridine diphosphoglucose (UDPG) of 5-FU-induced myelosuppression [23-26] and gastrointestinal toxicity [26, 27] without tumour protection supports this hypothesis. UDPG [27] administration enabled a 1.5-fold increase in 5-FU dosage; the better therapeutic index was most likely due to a prolonged TS inhibition, since UDPG decreased 5-FU incorporation into tumour RNA [28]. In addition, other murine studies demonstrate gastrointestinal toxicity due to RNA incorporation and not to increasing FdUMP levels [29]. Although 5-FU incorporation into RNA seems to be the major mechanism leading to 5-FU toxicity, inhibition of TS as an additional mechanism cannot be excluded since 5-FU administration can inhibit TS in mucosal tissues of mice and patients [30] and in bone marrow cells of rats and mice [30, 31]. The extent of TS inhibition is dependent on the administration route, with isolated liver perfusion resulting in the lowest TS inhibition [31]. In livers from mice, rats and patients, however, 5-FU did not inhibit TS but induced a 2-3fold increase in TS levels [30, 31]. We conclude that 5-FU toxicity is predominantly caused by 5-FU incorporation into RNA.

Uptake and clearance

In L-1210 cells [32] and in various human tumour cell lines and erythrocytes [33], FdUrd can be transported by a facilitated diffusion transport system (insensitive to nitrobenzylthioinosine). Rapid and not rate-limiting uptake of 5-FU occurs by facilitated transport [34]. Since negatively charged nucleotides cannot leave the cell, the active phosphorylated

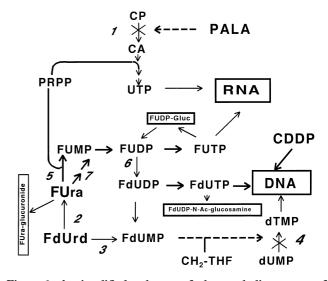


Figure 2. A simplified scheme of the anabolic routes of 5-FU and FdUrd activation and the interference with their modulators, N-phosponacetyl-L-aspartate (PALA), cisplatin (CDDP) and leucovorin (metabolised to 5, 10 methylenetetrahydrofolate, CH2-THF). CA, carbamyl aspartate; CP, carbamyl phosphate; UTP, uridine-5'-triphosphate; dTMP, 2'-deoxythymidine-5'-monophosphate; dUMP, 2'-deoxyuridine-5'-monophosphate; FUMP, 5-fluorouridine-5'-monophosphate; FdUMP, 5-fluoro-2'-deoxyuridine-5'-monophosphate; FUTP, 5-fluorouridine-5'-triphosphate; FdUTP, 5-fluoro-2'-deoxyuridine-5'-triphosphate; PRPP, 5-phosphoribosyl-1-pyrophosphate. Enzymes: 1, aspartate carbamoyl transferase (inhibited by PALA); 2, thymidine phosphorylase (TP); 3, thymidine kinase (TK); 4, thymidylate synthase (TS) (inhibited by FdUMP); 5, orotate phosphoribosyl transferase (OPRT); 6, ribonucleotide reductase; 7, sequential uridine phosphorylase-uridine kinase. CDDP forms DNA-platinum adducts.

fluoropyrimidine derivatives can accumulate in the cell [35]. In murine colon 26 tumours, rapid and extensive uptake occurs, allowing cellular 5-FU levels to be detected for up to 10 days [36, 37]. The elimination patterns of 5-FU after administration of 5-FU or FdUrd are similar in these studies, but 5-FU is cleared more rapidly from plasma resulting in a high tissue/plasma ratio (> 10) in patients [36].

Anabolism

After uptake, phosphorylation into the active nucleotides takes place (Figure 2). FdUrd can be phosphorylated directly by thymidine kinase (TK) to FdUMP, or cleaved by thymidine phosphorylase (TP) to 5-FU. In some circumstances, but not generally, FdUrd can act as a prodrug of 5-FU [22, 33, 38, 39]. FdUrd possibly has a dual mode of action. FdUMP can be phosphorylated to FdUDP and subsequently to FdUTP by pyrimidine nucleoside mono- and diphosphate kinases.

The predominant activation step for 5-FU is its phosphoribosylation to 5-fluorouridine-5'-monophosphate (FUMP) [40]. This reaction is catalysed by orotate phosphoribosyl transferase (OPRT) requiring the phosphate-donor 5-phosphoribosyl-1-pyrophosphate (PRPP) as a cosubstrate [41]. FUMP can also be formed in a two-step process via FUrd, catalysed by uridine phosphorylase, whose activity is elevated in proliferating tissues [40, 42], followed by uridine kinase. FUMP can be converted into FUDP by pyrimidine nucleoside monophosphate kinase [43]. FdUDP is then formed from FUDP by ribonucleotide reductase and can subsequently be phosphorylated by a non-specific pyrimidine nucleoside diphosphate kinase [44] to FdUTP using ATP as a cosubstrate [44].

TP is a bidirectional enzyme which theoretically can form FdUrd from 5-FU. However, in colorectal tumour tissues, the predominant direction of TP is toward formation of 5-FU from FdUrd due to lack of sufficient levels of the deoxyribose 1-phosphate donor [40,45]. Direct conversion of FdUrd to 5-FU is suggested in murine studies, where 5-FU after rapid FdUrd injection was present in tumour tissue for a longer period and in a higher concentration than in plasma [12,37]. These differences between whole body (plasma) and tumour 5-FU kinetics might be explained by tissue-specific isoenzymes of TP in mice and humans [46].

Catabolism

The active metabolites of both fluoropyrimidines can be converted back to 5-FU by several enzymes including 5'nucleotidases, phosphatases and phosphorylases (including TP) [47-49]. Free 5-FU itself can be degraded by dihydropyrimidine dehydrogenase (DPD), the first enzyme in the catabolic pathway. Reduction by DPD results in dihydrofluorouracil formation, followed by further enzymatic steps to β-ureidoproprionate (by dihydropyrimidinase) and to fluoroβ-alanine, CO₂ and NH₄⁺ (by ureidoproprionase). DPD is believed to play a role in sensitivity to 5-FU [50, 55], since congenital deficiency or low activity of DPD has been associated with enhanced toxicity and an increase in plasma 5-FU levels [52–54]. However, in populations of patients without this deficiency, clinical evidence of a relationship between DPD activity and response to treatment with 5-FU and cisplatin (CDDP) has not been found [55]. Thus, the importance of DPD may be limited to a relationship with toxicity, but not directly with antitumour activity.

MECHANISMS OF ACTION

Thymidylate synthase inhibition

FdUMP inhibits TS, the key enzyme in de novo pyrimidine synthesis [7, 18, 56, 57]. This inhibition can be augmented by co-administration of LV, which is the precursor of the TS co-substrate 5, 10-methylene-tetrahydrofolate (CH₂-THF). When adequate amounts of CH₂-THF are present, TS, FdUMP and CH₂-THF form a stable ternary complex [58– 62], which markedly increases the extent and duration of cellular TS inhibition, thereby enhancing antitumour activity. If CH₂-THF is present in inadequate amounts, FdUMP can dissociate from the enzyme allowing it to function. The kinetics of this process have been well described [59]. In patients, most pronounced inhibition was observed immediately after 5-FU administration (median residual TS activity of 46, 65 and 74% of total TS after 2, 23 and 45 h, respectively); with LV, TS activity was still 49% after 45 h [17, 63], demonstrating the essential role of reduced folates in maintaining enzyme inhibition in patients.

Incorporation into RNA

FUTP acts as a substrate for RNA polymerase and thus will be incorporated into mainly nuclear RNA [20,64,65]. Interference with the maturation of nuclear RNA might be related to cytotoxicity [7,20,65]. After injection of a therapeutic bolus dose of 5-FU, fluorinated nucleotides have been found in the RNA of tumours in animals for at least 1 week [37] and for 3 days in tumours of patients [66]. No relationship between 5-FU incorporation into RNA and response to 5-FU treatment has been observed in patients [63], while in the same group of 40 patients low TS levels and a high TS inhibition were related to response and high TS levels with no response [17,63].

Incorporation into DNA

FdUTP acts as a substrate for DNA polymerase and can be incorporated into DNA. However, after 5-FU treatment, DNA incorporation is only 10% of RNA incorporation [67–70] and is not considered an important mechanism of 5-FU cytotoxicity. In contrast, after FdUrd administration to CHO-K1 cells, equal percentages of incorporation into DNA and RNA are achieved with approximately 4-fold more incorporation into DNA after FdUrd than after 5-FU [70]. Furthermore, DNA single- and double-strand breaks occur in several cell lines after fluoropyrimidine treatment and are considered important determinants of FdUrd cytotoxicity but not of 5-FU [68,71]. Hence, it is feasible that DNA incorporation may contribute to the cytotoxicity of FdUrd, but not of 5-FU. However, the contribution of DNA incorporation to the overall effects of FdUrd has not yet been properly established.

Formation of sugar derivatives of 5-FU

FUDP sugars can be formed after 5-FU treatment [15,16,72–75] in various types of cells, serving as alternative substrates for the enzymes of UDP sugar metabolism [47]. FdUDP-N-acetylglucosamine and 5-FU glucuronide have been detected in human lymphoid cells [76] and rat hepatocytes [15,49,77], respectively. These sugars might serve as enzyme substrates, resulting in prolonged cell exposure to fluorinated nucleotides [15,47], but may also affect glycosylation reactions in tumour cells [15,78]. However, formation of sugars is not considered to play a significant role in the cellular effects of fluoropyrimidines.

PRECLINICAL ANTITUMOUR ACTIVITY IN COLORECTAL CANCER

In vitro studies

Growth inhibition and modulation of FdUrd and 5-FU by LV. Table 1 summarises the results of five studies [16, 79– 82] which compared the two fluoropyrimidines with and without LV in a variety of colorectal cancer cell lines. When given alone, FdUrd was more cytotoxic than 5-FU as expressed by the IC50 (molar concentration causing 50% growth inhibition) in a wide range of tumour cell lines. After at least 72 h incubation, the IC₅₀s of 5-FU ranged from 0.8 to 950 µM (data not shown) and FdUrd ranged from 0.36 nM to 3400 µM. In all but three cell lines, FdUrd was more cytotoxic than 5-FU on a molar basis. The dose-effect curve for each drug was sigmoidal. The modulation of FdUrd by LV was more pronounced, with median factors of modulation at their IC50s of 1.5 for 5-FU and 2.6 for FdUrd (Table 1). In most (88%) of these cell lines, FdUrd showed greater potentiation by LV than 5-FU. Furthermore, the cytotoxicity of FdUrd was enhanced by LV (potentiation factor > 1.5) in 17 out of 24 cell lines, compared with only 9 out of 24 cell lines for 5-FU. Although less pronounced, the same phenomenon was observed in a panel of gastric cancer cell lines [80]. In non-small cell lung cancer cells, neither fluoropyrimidine was modulated by LV in these cell lines which were resistant to fluoropyrimidines compared with colon cancer cell lines [80]. The difference between 5-FU and FdUrd may be due to the observation that in cell lines 5-FU can have different mechanisms of effect (RNA and DNA incorporation, or TS inhibition), which may not all be affected by LV. TS inhibition after FdUrd seems to be more general. Mechanistic differences may explain why modulation is different between the various tumour types.

Duration of drug exposure. In accordance with their cell cycle specificity, prolonged exposure of human cell lines to 5-FU or FdUrd greatly enhances growth inhibition [18,71,83,89]. In HCT-8 cells, a 24-fold increase in exposure time to FdUrd from 3 to 72 h led to a 38-fold reduction in IC₅₀ [83]. Remarkably, the modulation factor of LV in this study significantly decreased from 3.8 after 3 h to 2.0 after 72 h. In contrast, Moran and associates [79] found that prolonged exposure of WiDR cells with 5-FU led to increased enhancement of its cytotoxicity by LV [79], which was also observed by us in SW948 cells (Van der Wilt and colleagues, University Hospital VU, Amsterdam).

In summary, many *in vitro* studies show that growth inhibition due to FdUrd is greater than 5-FU. The difference is even more pronounced when LV is added, suggesting a significant role for TS in the cytotoxic effects of FdUrd. With a shorter exposure to FdUrd, LV does enhance the formation and stability of a ternary complex, thus prolonging inhibition of the catalytic activity of TS. In contrast, when the duration of FdUrd exposure is prolonged, the influence of LV modulation is decreased [83]. Apparently the binary complex between FdUMP and TS remains relatively stable during prolonged exposure to FdUrd so that addition of LV causes no enhancement of inhibition of TS catalytic activity. For 5-FU, addition of LV may cause a shift in its mechanism of

Table 1. In vitro cytotoxicity of 5-fluorouracil and 5-fluoro-2'-deoxyuridine alone and with LV modulation in human colorectal cancer tumour cells

	Drug exposure (days)	IC ₅₀ FdUrd (nM)		Potentiation by LV		
Cell line			IC₅₀ ratio*	FdUrd†	FUra†	- Reference
WiDR	3	4.0	625	8.0	2.5	79
WiDR	9–20	12.5	134	3.1	1.5	80
DLD-1	9–20	129.5	37	3.5	0.8	80
LOVO	9–20	34.4	96	3.9	1.5	80
COLO201	9–20	116	32	5.4	3.1	80
COLO320DM	9–20	80.5	12	4.6	1.4	80
COLO320	6	27.9	344	5.3	1.3	81
SW948	6	0.67	491	2.4	2.3	81
NCI-H548	4	3.4×10^6	0.17	1.1	1.0	82
NCI-H630	4	$17 imes 10^4$	0.35	1.6	1.3	82
NCI-H684	4	2.6×10^3	13	1.3	1.1	82
NCI-H508	4	1.1	150	1.5	1.5	82
NCI-H747	4	1.3×10^3	11	2.0	1.4	82
NCI-H716	4	4.7	5000	2.0	1.6	82
NCI-H498	4	38.4	143	2.2	2.7	82
SNU-C1	4	0.36	1333	1.7	1.8	82
SNU-C2a	4	12	11	1.6	1.6	82
SNU-C4	4	0.65	2545	2.1	1.7	82
SNU-C5	4	133	113	1.9	1.6	82
WiDR	3	3100	1.6	1.0	1.5	16
WiDR/F	3	2200	2.7	0.7	0.6	16
C26-10 (m)	3	4	133	1.3	0.9	16
C26-10/F (m)	3	15	43	2.5	0.9	16
CC 531 (r)	3	5200	0.35	0.9	1.4	16

In vitro cytotoxicity studies [79–82] in well-established human colorectal cancer cell lines comparing fluoropyrimidines and modulation by LV. *Ratio of the IC₅₀ of 5-FU over FdUrd (on a molar basis). †Ratio of the IC₅₀s of each fluoropyrimidine without and with leucovorin (LV). LV was given simultaneously with the fluoropyrimidines at a concentration of $10-20\,\mu\text{M}$ (except by Sinnige [81] who varied LV from $1-100\,\mu\text{M}$). The mean potentiation ratios for FdUrd and 5-FU are 2.6 ± 1.7 and 1.5 ± 0.6 , respectively (P<0.05, Student's *t*-test for unpaired data) which indicate overall greater potentiation of FdUrd by LV than 5-FU by LV.

action from RNA incorporation to TS inhibition, resulting in enhanced growth inhibition in several cell lines. Under conditions of metabolic modulations by LV, drug incorporation into cellular RNA is not altered significantly, neither in cell lines nor in tumour samples from mice [28] and patients [63]. These observations suggest that the cytotoxic effect of 5-FU can be optimised through LV modulation by maintenance of TS inhibition, while cancer cells treated with FdUrd and LV do not require prolonged exposure to achieve high levels of growth inhibition.

In vivo studies

In the last two decades many studies comparing 5-FU with FdUrd in various animal tumour models yielded insight into the relative effects of each drug alone, the probable best schedule for each drug in human studies and the level of benefit to be expected from modulators.

Comparison of FdUrd with 5-FU. Early in vivo experiments comparing FdUrd with 5-FU were performed by Corbett and associates in 1977 [85]. In various schedules (bolus administrations for 5 days, or weekly for 2-4 weeks) and different colon tumours (Colon 26, -36, -38 and -51), FdUrd yielded better therapeutic efficacy and more tumour-free survivors than 5-FU (6/69 versus 2/90, respectively). The highest responses for both drugs were seen in Colon 38. Studies by our group using various schedules (5 days continuous infusions, 5 days i.v. bolus, 3 weekly one hour infusions or 3 weekly i.v. bolus) in mice with Colon 26-B carcinoma revealed better antitumour activity of FdUrd compared with 5-FU at both equimolar and maximum tolerable doses. The most effective treatment schedule of FdUrd was a weekly i.v. bolus injection for 3 weeks. This schedule dependency is not observed with 5-FU treatment [12]. FdUrd is also more effective than 5-FU in rat models. The most effective schedule was also weekly high-dose administration [83]. Bartowski [86] studied rats bearing Novikoff hepatoma treated with either 5-day continuous infusion or five daily bolus injections. FdUrd had better antitumour activity than 5-FU at equimolar doses. With FdUrd the best antitumour activity was seen after continuous infusions. However, the maximum tolerated dose (MTD) was not determined and a weekly schedule was not studied. Continuous intra-arterial administration was the only route and schedule to produce significant antitumor activity for 5-FU in this study. In a similar experiment, rats bearing subcutaneous Novikoff hepatoma were given FdUrd via the femoral artery [87]. Five out of 6 rats (83%) had responses with five daily bolus administrations compared with 50% with equimolar doses given by continuous

Human colorectal xenograft tumours ($HxELC_2$ and $HxHC_1$) in immune-deprived mice showed better antitumour activity for one bolus injection of FdUrd compared with an equitoxic 5-FU dose [88]. Murine studies with Colon 26 by Iigo and associates [89] showed similar activity of 5-FU and FdUrd when given once i.p. After weekly therapy, superior antitumour activity of 5-FU compared with FdUrd was seen in the murine Colon 38 tumours [90]. This favourable antitumour activity of 5-FU compared with FdUrd as well as the antitumour activity of 5-FU differ from the results of other groups and might therefore not be representative [85, 91].

These findings show that intermittent FdUrd bolus administration is the best schedule of administration in these tumour models, and is better than any schedule of 5-FU.

Biochemical modulation. Extensive scheduling and dosefinding studies with LV revealed potentiation of the antitumour activity of 5-FU by LV given before and during 5-FU therapy in murine Colon 26-A and Colon 38 [60]. The Colon 38 tumour has lower TS activity, which may account for its greater sensitivity [61]. In both tumours the significant effects of LV on residual TS activity only occurred after 3 weeks of weekly administration [61]. First, inhibition of TS induced by LV plus 5-FU was more pronounced than 5-FU alone; second, LV prevented the 5-FU-induced increase in TS, which is seen in mice treated with 5-FU alone. Similarly, in another (Colon 26-B) model, superior FdUrd antitumour activity compared with 5-FU was related to more prolonged TS inhibition [37]. In another study, measurements of TS limited to 1 day in the Colon 38 tumour model of Iigo and associates [90] were too incomplete to correlate the antitumour activity of fluoropyrimidines with the extent of TS inhibition. In measurements of such short duration, LV is not likely to potentiate an almost complete TS inhibition. Although these in vivo data are limited, it might be concluded that prolonged TS inhibition and prevention of TS induction are of significant importance in the antitumour activity of fluoropyrimidines. The data between the various models are very different; when translated to the clinic, various different schedules may be used for different tumours.

Selective enhancement of the biochemical effects of fluoropyrimidines in tumours might improve therapeutic efficacy. Various strategies have been developed and evaluated by our group, using N-phosphonacetyl-L-aspartate (PALA) and CDDP as modulators [12, 91-94] in mice bearing Colon 26-A and B tumours. The antitumour activity of FdUrd was enhanced by PALA and even more so with a triple combination of PALA followed one day later by CDDP and FdUrd at an active dose [93]. This combination resulted in 66% complete responses (CR) compared to 25% with FdUrd alone and none with PALA + CDDP [12, 94]. The antitumour activity of 5-FU was enhanced by PALA from 10% partial responses (PR) to almost 60%, of which 10% were complete. Addition of a very low dose of CDDP resulted in 100% remissions of which 30% were complete [94]. Apparently triple PALA/fluoropyrimidine/CDDP combinations might be more effective than only two of these agents. The mechanism of action of PALA in this model appears to be through reduction in pyrimidine nucleotide pools, permitting unopposed fluoropyrimidine nucleotide action. The potentiation by PALA was related to the duration of decreased nucleotides [12]. Inhibition of TS activity by CDDP observed in Colon 26-A bearing mice treated with the chemoprotector WR-2721, 5-FU and modulating doses of CDDP might explain the synergism between fluoropyrimidines and platinum compounds [91].

Locoregional therapy. The aim of regional therapy is to improve the therapeutic index by delivering the highest concentrations of anticancer drugs directly to the site of the tumour achieving less systemic toxicity. Bartowski and associates [86] used rats bearing Novikoff hepatoma and showed that the highest tumour responses were found after administration of FdUrd directly into the hepatic artery compared with administration via the vena cava or portal vein. This finding is consistent with the known blood supply of liver metastases. Riemenschneider and associates [95] compared the effect of continuous infusions to intermittent locoregional therapy with FdUrd in the same animal system. Liver metastases treated by HAI from days 5 to 12 with

420 mg/kg FdUrd showed a low but significant tumour reduction. A bolus administration of 210 mg/kg on days 5 and 8 was ineffective. However, the schedule dependency of FdUrd was not taken into account as equal doses of FdUrd were used; much higher doses of FdUrd could have been used in the bolus arm of the study to achieve equal normal tissue toxicity.

In an attempt to overcome acquired resistance and minimise toxicity, Arisawa and colleagues used HAI FdUrd with or without mitomycin C in rats bearing K12/TRb liver metastases [96]. Treatment and resistance were evaluated by removal of the tumour after 28 days, and preparation of a single cell suspension followed by evaluation with a MTT assay. The antitumour activity of FdUrd with and without mitomycin C was comparable. However, decreased acquired resistance and toxicity were observed with 7 days of HAI FdUrd+one bolus dose of mitomycin C compared with 14 days of HAI FdUrd alone.

Summary of in vivo studies

These *in vivo* studies suggest that optimal FdUrd administration is with high-dose bolus injections. Furthermore, most studies show better therapeutic index for FdUrd compared to 5-FU. Also the use of modulators such as PALA, LV and CDDP leads to improvement of the therapeutic efficacy, while co-administration of mitomycin C might diminish acquired FdUrd resistance of hepatic tumours. Chronomodulation of continuous infusions can permit the delivery of a higher daily dose with less overall toxicity, due to circadian variation in TK and DPD [97–99]. The relative value of chronomodulation compared with intermittent bolus is unclear. However, in most circumstances, intermittent bolus injections are technically easier.

CLINICAL TRIALS WITH 5-FU OR FdUrd IN ADVANCED COLORECTAL CARCINOMA

Systemic FdUrd or 5-FU

In the 1960s, various groups compared i.v. FdUrd with 5-FU in patients with advanced colon cancer [99–102] (Table 2). Most studies showed no difference, but Ansfield and associates [99] showed significantly better response rates for FdUrd, possibly because of prolonged duration of treatment. In this study patients were treated for 11 consecutive days every 4 weeks, whereas in the other studies patients had shorter treatment periods [100–102] (Table 2).

Moertel and colleagues [103] compared rapid intravenous injection of FdUrd with continuous intravenous therapy in 128 patients. They concluded that 18-fold greater dose intensity could be achieved with bolus treatment for the same

toxicity level. In addition, significantly more objective regressions were found in the rapid injection group (17.5 versus 6.2%, respectively). Unfortunately, confirmatory studies have not been performed and the importance of this trial in considering the optimal administration schedule of FdUrd has been overlooked. Due to lower toxicity and costs, systemic 5-FU therapy has been preferred over FdUrd therapy [47]. Studies of the comparative efficacy and toxicity of FdUrd and 5-FU in large randomised clinical trials using current evaluation criteria are needed.

Hepatic arterial infusion

Ensminger and colleagues [6] showed that FdUrd was almost completely extracted (94–99%) by the liver compared to the 19–51% extraction of 5-FU, which is still much better than many other anticancer drugs [3,6]. Thus, FdUrd is ideally and 5-FU moderately suited to the pharmacological principle underlying HAI, in that the therapeutic index may be more favourable for those drugs by this route compared with systemic delivery. This hypothesis was tested in only one study in the late 1970s. HAI of 5-FU was compared with systemic administration of 5-FU in 61 patients with liver involvement of colorectal cancer. No significant differences in tumours responding were seen [104].

HAI of FdUrd has been evaluated in many clinical studies of advanced colorectal cancer [105-112], although not all studies were randomised [105]. In general, response rates have been superior to any systemic therapy, but possible survival benefits have been obscured by the cross-over of treatments. The median response rates in 10 studies of HAI of FdUrd was 45%, yielding a median survival of 17 months [3]. Three of these studies are shown in Table 3 [106–108], where HAI of FdUrd was compared with systemic FdUrd therapy in advanced colorectal cancer. In two of these studies, the response rate and survival were better for HAI [106, 107]. FdUrd has demonstrated its superiority over systemic 5-FU therapy in terms of response rates in four studies (Table 3; [109-112]). The observations can, however, be criticised, since in most of these trials cross-over was allowed and groups were too small to allow meaningful statistical evaluation [3, 9, 10, 13, 113, 114]. However, if the cross-over effects of the other trials were taken into account, HAI treatment was significantly better [3].

The most instructive studies are the French [110] and English [111] studies, where no cross-over was allowed, and in both trials HAI FdUrd resulted in significantly better median survival than systemic 5-FU therapy alone. Control groups were only treated according to the physician's preference and the HAI group was also treated with systemic

		Responses (%)			
Reference	5-FU†	FdUrd	No. patients	5-FU	FdUrd
Ansfield [99]	15 × 5→7.5 × 11	30 × 5→15 × 11	198	17	44*
Reitemeier [100]	$15 \times 5 \rightarrow 7.5 \times 4$	$40 \times 5 \rightarrow 20 \times 4$	168	12	23
ECGSTC† [102]	$15 \times 4 { ightarrow} 7.5 imes 4$	$30 \times 4 \rightarrow 15 \times 4$	104	27	4
Young [101]	$15 \times 4 \rightarrow 7.5 \times \text{tox}$ ‡	$30 \times 4 \rightarrow 15 \times tox$	86	15	16

Table 2. Comparison of systemic 5-FU with FdUrd

Early trials [99–102] comparing systemic 5-fluorouracil (5-FU) with 5-fluoro-2'-deoxyuridine (FdUrd) in patients with advanced colorectal cancer. *Drugs were given in mg/kg/d for 4 or 5 days, followed by half the initial dose every other day. Courses were repeated after 4–6 weeks until progression occurred. †Eastern Cooperative Group in Solid Tumor Chemotherapy. ‡Treatment until toxicity occurred. Significant difference (P<0.001) in response rates between 5-FU and FdUrd treatment was seen in one study [99].

Reference	No. of patients	Therapy		Partial responses %		Median survival (months)	
		HAI	Syst	HAI	Syst	HAI	Syst
Grage [104]	61	5-FU	5-FU	34	23	15.4	13.5
Chang [106]	64	FdUrd	FdUrd	62	17	17	12
Kemeny [107]	162	FdUrd	FdUrd	52	20	17	12
Hohn [108]	143	FdUrd	FdUrd	42	10	16.7	16.1
Martin [109]	74	FdUrd	5-FU	54	21	12.6	10.5
Rougier [110]	163	FdUrd	5-FU	43	9	15	11*
Allen-Mersh [111]	100	FdUrd	5-FU	50	0	13.5	7.2*
Wagman [112]	41	FdUrd	5-FU	56	0	_	_

Table 3. Comparison of systemic therapy with hepatic artery infusions of fluoropyrimidines

Randomised studies [104, 106–112] comparing systemic (Syst) fluoropyrimidines with hepatic artery infusions (HAI) of 5-fluorouracil (5-FU) or 5-fluoro-2'-deoxyuridine (FdUrd) in patients with liver metastases of colorectal cancer. Responses were significantly different in all studies except that of Grage [104]. *Significant (P<0.05) difference in survival or quality of life between systemic and HAI therapy was observed in two studies. In all but those two studies [110, 111], cross-over was allowed.

FdUrd therapy when extrahepatic disease occurred. The English group [111] was the only group to include quality of life assessments, and demonstrated improved quality of life for patients randomised to receive HAI FdUrd or best supportive care.

Since significant first-pass hepatic extraction of 5-FU occurs, a phase II study of HAI of 5-FU was conducted in Amsterdam in 37 patients. The overall response rate was 37.5%, which is in the same range as HAI FdUrd. As seen with HAI of FdUrd, no toxic side-effects were observed and patients responding to HAI 5-FU showed better inhibition of tumour TS than non-responders [17, 63]; no difference was observed for 5-FU incorporation into RNA. 5-FU did not inhibit liver TS, but in contrast induced a significant 2-3-fold increase in TS levels [30]. HAI FdUrd has not yet been compared with current optimal systemic therapy (5-FU+LV) in a randomised controlled trial. However, if the outcome of six randomised trials of systemic 5-FU and LV therapy is compared with HAI FdUrd in four other randomised trials [106-108, 110], HAI of FdUrd seems at least similar and possibly better in terms of objective responses. Furthermore, 1 and 2 year survival rates appear much better for FdUrd (Table 4) which may represent greater efficacy [3].

HAI FdUrd is accompanied by local toxicity, such as sclerosing cholangitis in 5-29% and chemical hepatitis [106–108, 110, 118]. In an attempt to reduce these side-effects, Kemeny and associates [115] compared dexamethasone and FdUrd to FdUrd alone in a randomised controlled trial. A remarkable increase in response rate was unexpectedly observed (71% for FdUrd + dexamethasone versus 40% for FdUrd alone, P=0.03, with a trend towards increased survi-

Table 4. Comparison of systemic 5-FU+LV with hepatic arterial infusion of FdUrd

		No. of	Partial responses (%)	Survival (%)	
Administration	Therapy	patients		1 year	2 year
Systemic HAI	5-FU+LV FdUrd	966 532	38 51	52 66	19 30

Randomised trials (total of 6 derived from [3]) using systemic 5-fluorouracil (5-FU)+leucovorin compared with four randomised studies [106–108, 110] using hepatic artery infusions (HAI) with 5-fluoro-2'-deoxyuridine (FdUrd) in patients with liver metastases of colorectal cancer, deduced from a review by Kemeny [3].

val; 23 months versus 15 months). In addition there was a significant reduction in local toxicity. The mechanism of this modulation remains unclear, but speculation includes protection of liver and endothelial cells by dexamethasone, resulting in the ability to prolong FdUrd therapy because of reduced toxicity.

Biochemical modulation of FdUrd may be beneficial, similarly to 5-FU modulation. A phase II study by Levin and Gordon [116], using a schedule of 6 days infusional LV and bolus FdUrd on days 2–6, showed a high response rate (55%) and high 1- and 2-year survival (73 and 50%, respectively) in previously untreated patients with advanced colorectal cancer, with a low incidence of severe toxicity. Modulation of FdUrd by LV using HAI in hepatic disease produced a median survival of 28.8 months [117]. In this study, high 1, 2, 3, 4 and even 5-year survival rates of 86, 62, 31, 15 and 7% were seen [117]. Dexamethasone was added to this schedule in a phase II study, producing similar survival rates and lower toxicity [118]. Other attempts to increase the therapeutic efficacy include alternating schedules with HAI FdUrd and systemic 5-FU, with beneficial therapeutic efficacy (minimal toxicity and similar survival rates of > 22 months) [119, 120]. These studies suggest that biochemical modulation of FdUrd by LV may be superior to FdUrd alone or to 5-FU+LV, but the strategy needs to be tested in randomised controlled comparative trials. The explanations for additional efficacy of dexamethasone with FdUrd but not with FdUrd + LV is unclear.

An argument against HAI FdUrd is the initial cost (financial costs, as well as the necessity for surgery for catheter and pump or port insertion). However, the overall expenses of HAI therapy are comparable to systemic 5-FU and LV administration after 1 year of treatment [3]. Since the median survival of patients treated with HAI exceeds this limit, HAI therapy cannot be dismissed on the basis of greater expense and the data strongly suggest that it provides better outcomes for patients with liver metastases.

Alternative approaches

Therapeutic 'synergism' between anticancer drugs has been considered worthwhile, whereby a mechanistic interaction may allow greater cytotoxicity in tumour tissues than the sum of the effects of each drug alone, but without greater toxicity in host tissue. In this regard, fluoropyrimidines have been combined with other anticancer agents. Patt and

colleagues [121] used HAI FdUrd + CDDP in patients with liver metastases from colorectal cancer, resulting in 52% response rates in 29 patients. In attempting to optimise total drug administration by giving high-dose i.v. bolus FdUrd with LV, an MTD of 1.65 g/m² FdUrd was achieved with 500 mg/m² LV in 2 h [22]. Alternatively, FdUrd may be given in the peritoneal cavity which might act as a drug reservoir and achieve prolonged low plasma concentrations reducing the toxicity. Doses of 3 g for three consecutive days could be achieved without toxicity [122]. A number of promising new drugs (e.g. campothecins and specific TS inhibitors) are currently under investigation [123, 124]. A recent randomised study of Tomudex compared to 5-FU-LV (low-dose LV daily × 5 schedule) revealed a 20% response rate for Tomudex compared to 13% for 5-FU-LV therapy [125]. However, the responses for FUra-LV are lower than in the original Mayo Clinic study (43%) [126]. CPT-11 data from randomised studies are awaited. More randomised controlled trials will be needed to demonstrate the benefit of such alternative approaches for patients with colorectal cancer.

Chronomodulation allows higher doses of FdUrd with decreased toxicity [14]. Chronotherapy of HAI FdUrd results in a 30% decrease in toxicity compared with flat-rate infusions [127], but is not yet accepted as a viable alternative to flat-rate infusions or bolus therapy and needs to be studied in large controlled and randomised trials.

CONCLUSIONS

Although much effort has been undertaken to optimise fluoropyrimidine therapy with either 5-FU or FdUrd for patients with advanced colorectal cancer, metastatic disease is still incurable. Although advances have been made, up to now the current treatment of these patients remains palliative. The most important advances, concerning improvement of quality of life and increased survival, are seen when metastases are limited to the liver and locoregional therapy can be administered using hepatic arterial infusions of FdUrd. If extrahepatic disease occurs, locoregional treatment can be combined with systemic 5-FU administration. Promising results are seen when FdUrd is modulated with either dexamethasone or LV and optimisation of the schedule might be reached with chronomodulated therapy.

The use of FdUrd in the treatment of patients with colorectal cancer might be more important than so far has been appreciated. The most effective schedule has yet to be clearly determined. Preclinical studies show different antitumour effects of 5-FU and FdUrd in various tumour model systems and schedules of administration, suggesting that different mechanisms of action may predominate in different circumstances for both fluoropyrimidines. In addition, recent research into the cellular pharmacology of FdUrd and the clinical observations that patients with metastatic colorectal cancer might benefit only from intermittent high bolus dose FdUrd rather than continuous infusions suggest that the optimal clinical administration of this drug may not be by continuous infusion, as had been thought previously. The favourable antitumour activity for systemic bolus FdUrd compared with 5-FU in the murine Colon 26 tumour model system [12], higher in vitro activity of FdUrd than 5-FU and reduced acquired resistance when treated for a shorter period with combination therapy [96] suggest that these schedules should be more intensively studied in the clinic. Modulating agents such as mitomycin C, platinum compounds, PALA or LV might be added to improve the outcome of colorectal cancer chemotherapy. When hepatic and extrahepatic disease co-exist, HAI FdUrd combined with systemic 5-FU therapy might improve survival. CPT-11 and Tomudex may not only be important new alternatives for fluoropyrimidine therapy [123–125], but should be evaluated in combination with fluoropyrimidines. Further clinical studies of the utility of fluoropyrimidines are warranted, and may ultimately show that in a specific schedule with or without biochemical modulators, FdUrd has superior efficacy to 5-FU in the management of patients with advanced colorectal cancer. Moreover, optimisation of fluoropyrimidine therapy may prove just as valuable as any of the promising new agents.

- Moller Jenson O, Esteve J, Renard H. Cancer in the European community and its member states. Eur J Cancer 1990, 26, 1167–1265.
- Weiss L, Grundmann E, Torhorst J. Haematogenous metastatic patterns in colonic carcinoma: an analysis of 1541 necropsies. 7 Pathol 1986, 150, 195–203.
- Kemeny N. Is hepatic infusion of chemotherapy effective treatment for liver metastases? Yes! *Important Adv Oncol* 1992a, 207–227
- 4. Heidelberger C, Chaudhuri NK, Danneberg PB, et al. Fluorinated pyrimidines, a new class of tumor-inhibitory compounds. *Nature*, 1957, **179**, 663–666.
- Bosch L, Harbers E, Heidelberger C. Studies on fluorinated pyrimidines V. Effects on nucleic acid metabolism in vitro. Cancer Res 1958, 18, 335–343.
- Ensminger WD, Rosowsky A, Raso V, et al. A clinical-pharmacological evaluation of hepatic arterial infusions of 5-fluoro-2'-deoxyuridine and 5-fluorouracil. Cancer Res 1978, 38 3784–3792.
- 7. Pinedo HM, Peters GJ. 5-Fluorouracil: biochemistry and pharmacology. *J Clin Oncol* 1988, **6**, 1653–1664.
- Peters GJ, Van Groeningen CJ. Clinical relevance of biochemical modulation of 5-fluorouracil. *Ann Oncol* 1991, 2, 469–480.
- De Takats PG, Kerr DJ, Poole CJ, Warren HW, McArdle CS. Hepatic arterial chemotherapy for metastatic colorectal carcinoma. Br 7 Cancer 1994, 69, 372–378.
- Patt YZ. Regional hepatic arterial chemotherapy for colorectal cancer metastatic to the liver: the controversy continues [editorial comment]. J Clin Oncol 1993, 11, 815–819.
- Levi F, Giacchetti S, Adam R, Zidani R, Metzger G, Misset J-L. Chronomodulation of chemotherapy against metastatic colorectal cancer. Eur J Cancer 1995, 31A, 1264–1270.
- Van Laar JAM, Durrani FA, Rustum YM. Antitumor activity
 of the weekly push schedule of 5-fluoro-2'-deoxyuridine ± Nphosphonacetyl-L-aspartate in mice bearing advanced colon
 carcinoma 26. Cancer Res 1993, 53, 1560–1564.
- Leichman CG. Prolonged infusion of fluorinated pyrimidines in gastrointestinal malignancies: a review of recent clinical trials. Cancer Invest 1994, 12, 166–175.
- 14. Von Roemeling R, Hrushesky WJM. Circadian patterning of continuous floxuridine infusion reduces toxicity and allows higher dose intensity in patients with widespread cancer. *J Clin Oncol* 1989, **11**, 1710–1719.
- 15. Peters GJ, Pinedo HM, Ferwerda W, De Graaf TW, Van Dijk W. Do antimetabolites interfere with the glycosylation of cellular glycoconjugates? *Eur J Cancer* 1990, **26**, 516–523.
- Van der Wilt CL, Peters GJ. New targets for pyrimidine antimetabolites for the treatment of solid tumors: I Thymidylate synthase. *Pharmacy World & Science* 1994, 16, 84–103.
- Peters GJ, Van der Wilt CL, Van Groeningen CJ, Smid K, Meijer S, Pinedo HM. Thymidylate synthase inhibition after administration of fluorouracil with or without leucovorin in colon cancer patients: implications for treatment with fluorouracil. J Clin Oncol 1994, 12, 2035–2042.
- Zhang ZG, Harstrick A, Rustum YM. Modulation of fluoropyrimidines: role of dose and schedule of leucovorin administration. *Semin Oncol* 1992, 19 (Suppl. 3), 10–15.
- Nord LD, Martin DS. Loss of murine tumor thymidine kinase activity in vivo following 5-fluorouracil (FUra) treatment by

- incorporation of FUra into RNA. Biochem Pharmacol 1991, 42, 2369-2375
- Spiegelman S, Sawyer RC, Nayak R, Ritzi E, Stolfi RL, Martin DS. Improving the antitumor activity of 5-fluorouracil by increasing its incorporation into RNA via metabolic modulation. *Proc Natl Acad Sci USA* 1980, 77, 4966–4970.
- Peters GJ, Van Groeningen CJ, Laurensse E, Pinedo HM. Thymidylate synthase from untreated human colorectal cancer and colonic mucosa: enzyme activity and inhibition by 5fluoro-2'-deoxyuridine-5'-monophosphate. *Eur J Cancer* 1991, 27A, 263–267.
- Creaven PJ, Rustum YM, Petrelli NJ, et al. Phase I and pharmacokinetic evaluation of floxuridine/leucovorin given on the Roswell Park weekly regimen. Cancer Chemother Pharmacol 1994, 34, 261–265.
- Van Groeningen CJ, Peters GJ, Leyva A, Laurensse E, Pinedo HM. Reversal of 5-fluorouracil-induced myelosuppression by prolonged administration of high-dose uridine. *J Natl Cancer Inst* 1989, 81, 157–162.
- Sawyer RC, Stolfi RL, Spiegelman S, Martin DS. Effect of uridine on the metabolism of 5-fluorouracil in the CD₈F₁ murine mammary carcinoma system. *Pharm Res* 1984, 2, 69–75.
- Peters GJ, Van Dijk J, Laurensse E, Van Groeningen CJ, Lankelma J, Leyva A, Nadal J, Pinedo HM. *In vitro* biochemical and *in vivo* biological studies of uridine "rescue" of 5-fluorouracil. *Br J Cancer* 1988, 57, 259–265.
- Bagrij T, Kralovansky J, Gyergyay F, Kiss E, Peters GJ. Influence of uridine treatment in mice on the protection of gastro-intestinal toxicity caused by 5-fluorouracil. *Anticancer Res* 1993, 13, 789–794.
- Codacci-Pisanelli G, Kralovansky J, Van der Wilt CL, et al. Modulation of 5-fluorouracil in mice using uridine diphosphoglucose. Clin Cancer Res 1997, 3, 309–315.
- 28. Codacci-Pisanelli G, Noordhuis P, Van der Wilt CL, et al. Incorporation of 5FU into RNA after high-dose 5FU treatment with uridine diphosphoglucoase (UDPG) rescue in mice is not related with antitumor activity, but thymidylate synthase (TS) inhibition. *Proc AACR* 1997, 38, 478 (Abstract 3201).
- Houghton JA, Houghton PJ, Wooten RS. Mechanism of induction of gastrointestinal toxicity in the mouse by 5-fluorouracil, 5-fluorouridine, and 5-fluoro-2'-deoxyuridine. *Cancer Res* 1979, 39, 2406–2413.
- 30. Van der Wilt CL, Van Groeningen CJ, Pinedo HM, et al. 5-Fluorouracil-leucovorin induced inhibition of thymidylate synthase in normal tissues of man and mouse. A role in toxicity? J Cancer Res Clin Oncol, in press.
- Van der Wilt CL, Marinelli A, Pinedo HM, et al. The effect of different routes of administration of 5-fluorouracil on thymidylate synthase inhibition in the rat. Eur J Cancer 1995, 31A, 754-760.
- Crawford CR, Ng CYC, Noel LD, Belt JA. Nucleoside transport in L1210 murine leukemia cells. Evidence for three transporters. *J Biol Chem* 1990, 265, 9732–9736.
- Uchida M, Ho DHW, Kamiya K, et al. Transport and intracellular metabolism of fluorinated pyrimidines in cultured cell lines. In Mikanagi K, Nishioka K, Kelley WN, eds. Purine and Pyrimidine Metabolism in Man VIB. New York, Plenum Press, 1989, 321–326.
- Wohlhueter RM, McIvor RS, Plageman PGW. Facilitated transport of uracil and 5-fluorouracil and permeation of orotic acid into cultured mammalian cells. J Cell Physiol 1980, 104, 309–319.
- 35. Keppler D, Holstege A. Pyrimidine nucleotide metabolism and its compartmentation. In Sies H, ed. *Metabolic Compartmentation*, London, Academic Press, 1982, 147–203.
- Peters GJ, Lankelma J, Kok RM, et al. Long retention of high concentrations of 5-fluorouracil in human and murine tumors compared to plasma. Cancer Chemother Pharmacol 1993, 31, 269-276.
- 37. Van Laar JAM, Van der Wilt CL, Smid K, et al. Therapeutic efficacy of fluoropyrimidines depends on the duration of thymidylate synthase inhibition in the murine Colon 26-b carcinoma tumor model. Clin Cancer Res 1996, 2, 1327–1333.
- Woodman PW, Sarrif AM, Heidelberger C. Specificity of pyrimidine nucleoside phosphorylases and the phosphorolysis of 5-fluoro-2'-deoxyuridine. *Cancer Res* 1980, 40, 507–511.

- Uchida M, Brown N, Ho DHW. Enzymatic conversion of 5fluoro-2'-deoxyuridine to 5-fluorouracil or 5-fluoro-2'-monophosphate in human tissues. *Anticancer Res* 1990, 10, 779–784.
- Peters GJ, Van Groeningen CJ, Laurensse EJ, Pinedo HM. A comparison of 5-fluorouracil metabolism in human colorectal cancer and colon mucosa. *Cancer* 1991, 68, 1903–1909.
- 41. Heidelberger C, Danenberg PV, Moran RG. Fluorinated pyrimidines and their nucleosides. *Adv Enzyme* 1983, **54**, 57–119.
- 42. Sköld O. Enzymes of uracil metabolism in tissues with different growth characteristics. *Biochim Biophys Acta* 1960, 44, 1–12.
- Hande KR, Chabner BA. Pyrimidine nucleoside monophosphate kinase from human leukemic blast cells. *Cancer Res* 1978, 38, 579–585.
- Agrawal RP, Parks RE. Erythrocytic nucleoside diphosphokinase. J Biol Chem 1971, 246, 2258–2264.
- Peters GJ, Laurensse E, Leyva A, Pinedo HM. Purine nucleosides as cell-specific modulators of 5-fluorouracil metabolism and toxicity. Eur J Cancer Clin Oncol 1987, 23, 1869–1881.
- 46. El Kouni MH, El Kouni MM, Naguib FN. Differences in activities and substrate specificity of human and murine pyrimidine nucleoside phosphorylases: implications for chemotherapy with 5-fluoropyrimidines. *Cancer Res* 1993, 53, 3687– 3693
- 47. Weckbecker G. Biochemical pharmacology and analysis of fluoropyrimidines alone and in combination with modulators. *Pharmacol Ther* 1991, **50**, 367–424.
- 48. Peters GJ, Laurensse E, Leyva A, Lankelma J, Pinedo HM. Sensitivity of human, murine and rat cells to 5-fluorouracil and 5'-deoxy-5-fluorouridine in relation to drug-metabolizing enzymes. *Cancer Res* 1986, 46, 20–28.
- 49. Sweeny DJ, Barnes S, Diasio RB. Formation of conjugates of 2-fluoro-β-alanine and bile acids during the metabolism of 5-fluorouracil and 5-fluoro-2'-deoxyuridine in the isolated perfused rat liver. Cancer Res 1988, 48, 2010–2014.
- Beck A, Etienne MC, Cheradame S, et al. A role for dihydropyrimidine dehydrogenase and thymidylate synthase in tumor sensitivity to fluorouracil. Eur J Cancer, 1994, 10, 1517–1522.
- Peters GJ, Van der Wilt CL, Van Groeningen CJ. Predictive value of thymidylate synthase and dihydropyrimidine dehydrogenase. Eur J Cancer 1994, 30A, 1408–1411.
- 52. Diasio RB, Harris BE. Clinical pharmacology of 5-fluorouracil. *Clin Pharmacokin* 1989, **16**, 215–237.
- 53. Fleming RA, Milano G, Thys A, et al. Correlation between dihydropyrimidine dehydrogenase activity in peripheral mononuclear cells and systemic clearance of 5-fluorouracil in cancer patients. Cancer Res 1992, 52, 2899–2902.
- Takimoto CH, Lu ZH, Zhang R, et al. Severe neurotoxicity following 5-fluorouracil-based chemotherapy in a patient with dihydropyrimidine dehydrogenase deficiency. Clin Cancer Res 1996, 2, 477–481.
- Etienne MC, Cheradame S, Fischel JL, et al. Response to fluorouracil therapy in cancer patients: the role of tumoral dihydropyrimidine dehydrogenase activity. J Clin Oncol 1995, 13, 1663–1670.
- 56. Rustum YM. Modulation of fluoropyrimidines by leucovorin: rationale and status. *J Surg Oncol Suppl* 1991, **2**, 116–123.
- Langenbach RJ, Danenberg PV, Heidelberger C. Thymidylate synthetase: mechanism of inhibition by 5-fluoro-2'-deoxyuridylate. *Biochem Biophys Res Commun* 1972, 48, 1565–1571.
- Santi DV, McHenry CS, Sommer H. Mechanism of interaction of thymidylate synthetase with 5-fluorodeoxyuridine. *Biochemistry* 1974, 13, 471–481.
- 59. Danenberg PV. Thymidylate synthetase—a target enzyme in cancer chemotherapy. *Biochim Biophys Acta* 1977, 473, 73–92.
- Nadal JC, Van Groeningen CJ, Pinedo HM, Peters GJ. Schedule-dependency of *in vivo* modulation of 5-fluorouracil by leucovorin and uridine rescue in murine colon carcinoma. *Invest New Drugs* 1989, 7, 163–172.
- 61. Van der Wilt CL, Pinedo HM, Smid K, Peters GJ. Elevation of thymidylate synthase following 5-fluorouracil treatment is prevented by the addition of leucovorin in murine colon tumors. *Cancer Res* 1992, **52**, 4922–4928.
- 62. Peters GJ, Hoekman K, Van Groeningen CJ, et al. Potentiation of 5-fluorouracil induced inhibition of thymidylate synthase in human colon tumors by leucovorin is dose dependent. In Ayling JE, Nair MG, Baugh CM, eds. Chemistry and Biology of

- Pteridines and Folates. New York, Plenum Press, 1993, 613-616.
- 63. Peters GJ, Van der Wilt CL, Van Groeningen CJ, et al. Effect of different leucovorin formulations on 5-fluorouracil induced thymidylate synthase inhibition in colon tumors and normal tissues from patients in relation to response to 5-fluorouracil. In Pfleiderer W & Rokos H, eds. Chemistry and Biology of Pteridines and Folates (Proc. 11th International Symposium). Blackwell Science, Berlin, 1997, 145–150.
- 64. Chaudhuri NK, Montag BJ, Heidelberger C. Studies on fluorinated pyrimidines. III. The metabolism of 5-fluorouracil-2C¹⁴ and 5-fluoroorotic-2-C¹⁴ acid in vivo. Cancer Res 1958, 18, 318–328.
- 65. Mandel HG. The target cell determinants of the antitumor actions of 5-FU: does FU incorporation into RNA play a role. *Cancer Treat Rep* 1981, **65** (Suppl. 3), 63–71.
- 66. Peters GJ, Noordhuis P, Komissarov A, *et al.* Quantification of 5-fluorouracil incorporation into RNA of human and murine tumors as measured with a sensitive gas-chromatography–mass spectometry assay. *Analytical Biochem* 1995, **231**, 157–163.
- 67. Tanaka M, Yoshida S, Saneyoshi M, Yanaguchi T. Utilization of 5-fluorouridine triphosphate and 5-fluoro-2'-deoxycitidine triphosphate in DNA synthesis by DNA polymerases α and β from calf thymus. *Cancer Res* 1981, 41, 4132–4135.
- Lönn U, Lönn S. DNA lesions in human neoplasmatic cells and cytotoxicity of 5-fluoropyrimidines. *Cancer Res* 1986, 46, 3866–3871.
- Aschele C, Sobrero A, Faderan MA, Bertino JR. Novel mechanism(s) of resistance to 5-fluorouracil in human colon cancer (HCT-8) sublines following exposure to two different clinically relevant dose schedules. *Cancer Res* 1992, 52, 1855– 1864.
- Willmore E, Durkacz BW. Cytotoxic mechanisms of 5-fluoropyrimidines. Relationships with poly(ADP-ribose) polymerase activity, DNA strand breakage and incorporation into nucleic acids. *Biochem Pharmacol* 1993, 46, 205–211.
- Yin MB, Rustum YM. Comparative DNA strand breakage induced by FUra and FdUrd in human ileocecal adenocarcinoma (HCT-8) cells: relevance to cell growth inhibition. *Cancer Communications* 1991, 3, 45–51.
- 72. Holstege A, Herrmann B, Keppler D. Increased formation of nucleotide derivatives of 5-fluorouridine in hepatoma cells treated with inhibitors of pyrimidine synthesis and D-galactosamine. *FEBS Lett* 1978, **95**, 361–365.
- Holstege A, Keppler D. Effects of p-glucosamine and 6azauridine on nucleotide contents, 5-fluorouridine uptake and cytotoxicity in TA3 mammary tumor cells. J Natl Cancer Inst 1986, 76, 485–492.
- 74. Anukarahanonta T, Holstege A, Keppler D. Selective enhancement of 5-fluorouridine uptake and action in rat hepatomas *in vivo* following pretreatment with D-galactosamine and 6-azauridine or N-(phosphonoacetyl)-L-aspartate. *Eur J Cancer* 1980, **16**, 1171–1180.
- Peters GJ, Laurensse E, Lankelma J, Leyva A, Pinedo HM. Separation of several 5-fluorouracil metabolites in various melanoma cell lines. Evidence for the synthesis of 5-fluorouracil nucleotide sugars. Eur J Cancer Clin Oncol 1984, 20, 1425–1431.
- Peterson MS, Ingraham HA, Goulian M. 2'-Deoxyribosyl analogues of UDP-N-acetylglucosamine in cells treated with methotrexate and 5-fluorodoxyuridine. *J Biol Chem* 1983, 258, 10831–10834.
- 77. Sommadossi J-P, Cross DS, Gewirtz DAA, Goldman ID, Cano J-P, Diasio RB. Evidence from rat hepatocytes of an unrecognized pathway of 5-fluorouracil metabolism with the formation of glucuronide derivative. *Cancer Res* 1985, 45, 2450–2455.
- 78. De Graaf TW, Slot SS, Peters GJ, Van Dijk W. Changes in the glycosylation of L1210 cells after exposure of various antimetabolites. *Eur J Cancer* 1993, **29A**, 1760–1765.
- 79. Moran RG, Scanlon KL. Schedule-dependent enhancement of the cytotoxicity of fluoropyrimidines to human carcinoma cells in the presence of folinic acid. *Cancer Res* 1991, **51**, 4618–4623.
- Sugimoto Y, Ohe Y, Nishio K, Ohmori T, Fujiwara Y, Saijo N.
 In vitro enhancement of fluoropyrimidine-induced cytotoxicity by leucovorin in colorectal and gastric carcinoma cell lines but not in non-small lung carcinoma cell lines. Cancer Chemother Pharmacol 1992, 30, 417–422.

- 81. Sinnige HAM, Timmer-Bosscha H, Peters GJ, De Vries EGE, Mulder NH. Combined modulation by leucovorin and α-2a interferon of fluoropyrimidine mediated growth inhibition. *Anticancer Res* 1993, 13, 1335–1340.
- Park J-G, Collins JM, Gazdar AD, et al. Enhancement of fluorinated pyrimidine-induced cytotoxicity by leucovorin in human colorectal carcinoma cell lines. J Natl Cancer Inst 1988, 80, 1560–1564.
- Cao S, Zhang Z, Creaven PJ, Rustum YM. 5-Fluoro-2'-deoxyuridine: Role of schedule in its therapeutic efficacy. In Rustum YM, ed. Novel Approaches to Selective Treatment of Human Solid Tumours: Laboratory and Clinical Correlation. New York, Plenum Press, 1993, 1–8.
- Van Ark-Otte J, Peters GJ, Pizao PE, Keepers YPAM, Giaccone G. In vitro schedule dependency of EO9 and miltefostine in comparison to standard drugs in colon cancer cells. Int J Oncol 1994, 4, 709–715.
- Corbett TH, Griswold DP, Roberts BJ, Peckham JC, Schabel FM. Evaluation of single agents and combinations of chemotherapeutic agents in mouse colon carcinomas. *Cancer* 1977, 40, 2660–2680.
- Bartowski R, Berger MR, Aguiar JLA, et al. Experiments on the efficacy and toxicity of locoregional chemotherapy of liver tumors with 5-fluoro-2'-deoxyuridine (FUDR) and 5-fluorouracil (5-FU) in an animal model. J Cancer Res Clin Oncol, 1986, 111, 42–46.
- 87. Naser-Hijazi B, Berger MR, Schmal D, Schlag P, Hull WE. Locoregional administration of 5-fluoro-2'-deoxyuridine (FdUrd) in Novikoff hepatoma in the rat: effects of dose and infusion time on tumor growth and on FdUrd metabolite levels in ¹⁹F-NMR spectroscopy. J Cancer Res Clin Oncol 1991, 117, 295–304.
- Houghton JA, Houghton PJ. On the mechanism of cytotoxicity of fluorinated pyrimidines in four human colon adenocarcinoma xenografts maintained in immune-deprived mice. *Cancer* 1980, 45, 1159–1167.
- Iigo M, Nishikata K-I, Nakajima Y, Araki E. Effects of antitumor agents on subcutaneous implants and hepatic metastases of Colon carcinoma 26 in mice. Jpn J Cancer Res 1992, 83, 397–401.
- Iigo M, Nishikata K-I, Hoshi A. In vivo antitumor effects of fluoropyrimidines on colon adenocarcinoma 38 and enhancement by leucovorin. Jpn J Cancer Res 1992, 83, 392–396.
- 91. Van der Wilt CL, Van Laar JAM, Gyergyay F, Smid K, Peters GJ. Biochemical modification of the toxicity and the antitumor effect of 5-fluorouracil and cisplatinum by WR-2721. *Eur J Cancer* 1992, **28A**, 2017–2014.
- 92. Van Laar JAM, Rustum YM, Van der Wilt CL, et al. Tumor size and origin determine the antitumor activity of cisplatin or 5-fluorouracil and its modulation by leucovorin in murine colon carcinoma. 1996, **39**, 79–89.
- 93. Van Laar JAM, Mayhew EG, Cao S, Durrani F, Peters GJ, Rustum YM. Modulation of the antitumor activity of cisplatin and/or 5-fluoro-2'-deoxyuridine by N-phosphonacetyl-L-aspartate in murine colon carcinoma #26. Eur J Cancer 1995, 31A, 974–976.
- 94. Durrani FA, Cao S, Van Laar JAM, Rustum YM. Modulation of the antitumor activity of 5-fluorouracil and cisplatinum by *N*-phosphonacetyl-L-aspartate in the murine colon carcinoma #26. *Int J Oncol* 1994, 5, 1065–1068.
- 95. Riemenschneider Th, Ruf C, Spath G, Stuhldreier G, Elmouaouy A. Continuous or bolus chemotherapy with 5-fluoro-2'-deoxyuridine in transplanted experimental tumors? *J Cancer Res Clin Oncol* 1988, 114, 482–486.
- Arisawa Y, Sutanto-Ward E, Dalton RD, Sigurdson ER. Short-term intrahepatic FUdR infusion combined with bolus mitomycin C: Reduced risk for developing drug resistance. J Surg Oncol 1994, 56, 75–80.
- Zhang R, Lu Z, Liu T, Soong SJ, Diasio RB. Relationship between circadian-dependent toxicity of 5-fluorodeoxyuridine and circadian rhythms of pyrimidine enzymes: possible relevance to fluoropyrimidine chemotherapy. *Cancer Res* 1993, 53, 2816–2822.
- 98. Kemeny MM, Alava G, Oliver JM. The effects on liver metastases of circadian patterned continuous hepatic artery infusion of FUDR. *H.P.B. Surg* 1994, 7, 219–224.

- 99. Ansfield FJ, Curreri AR. Further clinical comparison between 5-fluorouracil (5-FU) and 5-fluoro-2'-deoxyuridine (5-FUDR). *Cancer Chemother Rept* 1963, **32**, 101–105.
- 100. Reitemeier RJ, Moertel CG, Hahn RG. Comparison of 5-fluorouracil and 2'-deoxy-5-fluorouridine in treatment of patients with advanced adenocarcinoma of the colon or rectum. Cancer Chemother Rep 1965, 44, 39–43.
- 101. Young CW, Ellison RR, Sullivan RD, et al. The clinical evaluation of 5-fluorouracil and 5-fluoro-2'-deoxyuridine in solid tumors in adults. Cancer Chemother Rept 1960, 6, 17–20.
- 102. Eastern Cooperative Group in Solid Tumor Chemotherapy. Comparison of antimetabolites in the treatment of breast cancer and colon cancer. §AMA 1967, 200, 101–118.
- 103. Moertel CG, Reitemeier RJ, Hahn RG. A controlled comparison of 5-fluoro-2'-deoxyuridine therapy administered by rapid intravenous injection and by continuous intravenous therapy. *Cancer Res* 1976, 27, 549–552.
- 104. Grage TB, Vassilopoulus PP, Shingleton WW, et al. Results of a prospective randomized study of hepatic artery infusion with 5-fluorouracil versus intravenous 5-fluorouracil in patients with metastases from colorectal cancer: A Central Oncology Group study. Surgery 1979, 86, 550–555.
- 105. Weiss GR, Garnick MB, Osteen RT, et al. Long-term hepatic arterial infusion of 5-fluorodeoxyuridine for liver metastases using an implantable infusion pump. J Clin Oncol 1983, 1, 337– 345.
- 106. Chang AE, Schneider PD, Sugarbaker PH, Simpson C, Culnane M, Steinberg SM. A prospective randomized trial of regional versus systemic continuous 5-fluorodeoxyuridine chemotherapy in the treatment of colorectal liver metastases. *Ann Surg* 1987, 206, 685–693.
- 107. Kemeny N, Daly J, Reichman B, Geller N, Botet J, Oderman P. Randomized study of intrahepatic versus systemic infusion of fluorodeoxyuridine in patients with liver metastases from colorectal cancer. *Ann Int Med* 1987, 107, 459–465.
- 108. Hohn DC, Stagg RJ, Friedman MA, et al. A randomized trial of continuous intravenous versus hepatic intra-arterial floxuridine in patients with colorectal cancer metastases to the liver: The Northern California Oncology Group Trial. J Clin Oncol 1989, 7, 1646–1654.
- 109. Martin JK, O'Connell MJ, Wieand HS, et al. Intra-arterial floxuridine vs systemic fluorouracil for hepatic metastases from colorectal cancer. Arch Surg 1990, 125, 1022–1027.
- 110. Rougier P, Laplanche A, Huguier M, et al. Hepatic arterial infusion of floxuridine in patients with liver metastases from colorectal carcinoma: long-term results of a prospective randomized trial. J Clin Oncol 1992, 10, 1112–1118.
- Allen-Mersh TG, Earlam S, Fordy C, Abrams K, Houghton J. Quality of life and survival with continuous hepatic-artery floxuridine infusion for colorectal liver metastases. *Lancet* 1994, 344, 1255–1260.
- 112. Wagman LD, Kemeny MM, Leong L, Terz JJ. A prospective randomized evaluation of the treatment of colorectal cancer metastatic to the liver. J Clin Oncol 1990, 8, 1885–1893.
- 113. Begos DG, Ballantyne GH. Regional chemotherapy for colorectal liver metastases: thirty years without patient benefit. J Surg Oncol 1994, 56, 139–144.

- 114. O'Connell MJ. Is hepatic infusion of chemotherapy effective treatment for liver metastases? No! *Important Adv Oncol* 1992, 229–234.
- 115. Kemeny N, Seiter K, Niedzwiecki D, et al. A randomized trial of intrahepatic infusion of fluorodeoxyuridine with dexamethasone versus fluorodeoxyuridine alone in the treatment of metastatic colorectal cancer. Cancer 1992, 69, 327–334.
- Levin RD, Gordon JHTI. Fluorodeoxyuridine with continuous leucovorin infusion. A phase II clinical trial in patients with metastatic colorectal cancer. *Cancer* 1993, 72, 2895–2901.
- 117. Kemeny N, Seiter K, Conti JA, et al. Hepatic arterial floxuridine and leucovorin for unresectable liver metastases from colorectal carcinoma. New dose schedules and survival update. Cancer 1994, 73, 1134–1142.
- 118. Kemeny N, Conti JA, Blumgart L, et al. A phase II study of hepatic arterial infusion of FUDR, leucovorin and dexamethasone for unresectable liver metastases from colorectal carcinoma. J Clin Oncol 1994, 12, 2288–2295.
- 119. Stagg RJ, Venook AP, Chase JL, et al. Alternating hepatic intraarterial floxuridine and fluorouracil: a less toxic regimen for treatment of liver metastases from colorectal cancer. J Natl Cancer Inst 1991, 83, 423–428.
- 120. Seiter K, Kemeny N, Sigurdson E, Cohen A, Oderman P. A phase I trial of hepatic arterial fluorodeoxyuridine combined with systemic 5-fluorouracil for the treatment of metastases from colorectal cancer. Reg Cancer Treat 1991, 3, 293–297.
- 121. Patt YZ, Boddie AW, Charnsangavej, et al. Hepatic arterial infusion with floxuridine and cisplatin: overriding importance of antitumor effect versus degree of tumor burden as determinants of survival among patients with colorectal cancer. *J Clin Oncol* 1986, 4, 1356–1364.
- 122. Muggia FM, Chan KK, Russell C, et al. Phase I and pharmacologic evaluation of intraperitoneal 5 fluoro 2' deoxyuridine. Cancer Chemother Pharmacol 1991, 28, 241–250.
- 123. Armand JP, Ducreux M, Mahjoubi M, et al. CPT-11 (Irinote-can) in the treatment of colorectal cancer. Eur J Cancer 1995, 31A, 1283–1287.
- 124. Jackman AL, Farrugia DC, Gibson W, et al. ZD1696 (tomudex): a new thymidylate synthase inhibitor with activity in colorectal cancer. Eur J Cancer 1995, 31A, 12778–1282.
- 125. Cunningham D, Zalcberg JR, Rath U, et al. and the 'Tomudex' Colorectal Cancer Study Group. Tomudex (ZD1694): results of a randomised trial in advanced colorectal cancer demonstrate efficacy and reduced mucositis and leucopenia. Eur J Cancer 1995, 31A, 1945–1954.
- O'Connell MJ. A phase III trial of 5-fluorouracil and leucovorin in the treatment of advanced colorectal cancer; a Mayo-Clinic/ North Central Cancer Treatment Group study. *Cancer* 1989, 63, 1026–1030.
- 127. Wesen C, Hrushkey WJM, Von Roemeling RV. Circadian modification of intra-arterial 5-fluoro-2'-deoxyuridine infusion rate reduces its toxicity and permits higher dose intensity. J Infus Chemother 1992, 2, 69-75.

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